

Supplementary Table 1. Major antibiotic resistant bacteria, clinical manifestation, associated and available antibiotics for treatment (only examples are given).

Resistant Bacteria/Type/Region of High Prevalence	Symptoms/Representative Infections	Clinical	Antibiotics Associated with Resistance	Available Treatments	Ref
<i>Staphylococcus aureus</i> (S. aureus) (MRSA)/ gram-positive, cocci, Eastern Mediterranean Region (0-92%)	Skin and soft tissue infections, toxic shock syndrome, osteomyelitis pneumonia/lung infection, endocarditis, meningitis, urinary tract infection (UTI)		beta-lactam antibiotics (e.g., penicillin, oxacillin, monobactams, nafcillin, amoxicillin, and most cephalosporins) erythromycin	Tedizolid, tigecycline, vancomycin or daptomycin, ceftaroline, telavancin; linezolid; dalbavancin, oritavancin, clindamycin, minocycline, doxycycline, linezolid	[1-5]
<i>S. aureus</i> (VISA/hVISA/VRSA)/ gram-positive, cocci, South-East Asia Region (3.42–6.81%)	Skin and soft tissue infections, UTI, bacteraemia, pneumonia/ lung infection, osteomyelitis, endocarditis, meningitis		beta-lactam antibiotics, vancomycin, erythromycin	Minocycline, or quinupristin-dalfopristin, daptomycin, linezolid, telavancin, ceftaroline	[2,6-9]
<i>Streptococcus pyogenes</i> / gram positive, cocci, Europe region (4–40%)	Pharyngitis, necrotizing fasciitis, toxic shock syndrome, autoimmune diseases		macrolides, clindamycin, and lincosamide	Penicillin, cephalosporins, macrolides, and clindamycin	[10-14]
<i>Streptococcus pneumoniae</i> / gram-positive, diplococcus, African Region (1-100%)	Pneumonia/ lung infection, otitis media, sinusitis, bronchitis, bacteraemia, peritonitis, cellulitis, arthritis, meningitis		optochin, ceftriaxone, cefotaxime, penicillin, Cefuroxime, cephalosporins, erythromycin, tetracycline, doxycycline	Vancomycin +/- rifampin; gemifloxacin, moxifloxacin, levofloxacin) linezolid; clindamycin; imipenem/cilastatin, ceftriaxone plus	[2,5,15-17]
<i>Escherichia coli</i> - CTX-M extended spectrum beta-lactamases (ESBL)/ gram- negative, rod, South-East Asia Region 19-95% ()	UTI, diarrhoea, colitis		Penicillin, cephalosporins, fluoroquinolones, aztreonam	Meropenem, carbapenems (ertapenem, doripenem), imipenem, cilastatin, fosfomycin, trimethoprim-sulfamethoxazole, cefepime	[5,18-22]
<i>Enterococcus faecium</i> (VRE)/ gram- positive, oval cocci Latin America (0-48%)	Pelvic infections, endocarditis, neonatal infections and UTI		Vancomycin, gentamicin, ampicillin, beta lactams and aminoglycosides, streptomycin	Linezolid, daptomycin, streptogramins tigecycline, quinupristin-dalfopristin, sultamicillin	[23-30]
<i>Pseudomonas aeruginosa</i> (MDR)/gram-negative, rod Latin America (0-48%)	Skin and soft-tissue infections, UTIs, bloodstream infections, pneumonia/lung infection, endocarditis, meningitis		Imipenem/cilastatin, meropenem, non-antipseudomonal penicillins, cephalosporins	Colistin, ceftazidime-avibactam or ceftolozane-tazobactam, polymyxin B	[31-37]
<i>Klebsiella pneumoniae</i> (ESBL)/ gram- negative, rod South-East Asia Region (20-72%)	Bacteraemia, sepsis, pneumonia, UTIs, conjunctivitis, and surgical site infections		Beta-lactam, ciprofloxacin, cephalosporins; aztreonam; aminoglycosides and norfloxacin	Carbapenems, doripenem, flomoxef, Piperacillin ceftolozane-tazobactam, ceftazidime-avibactam	[5,38-42]

<i>Mycobacterium tuberculosis</i> (MDR-TB)/ acid-fast Indo-Pak Region (1.6-21.6%)	Tuberculosis (lung infection), osseous tuberculosis, night sweats, fever, weight loss	Isoniazid; rifampin; streptomycin	Pyrazinamide, a fluoroquinolone, an injectable agent (i.e., amikacin), ethionamide, cyclomerize, linezolid, clofazimine	[43-48]
<i>Acinetobacter baumannii</i> / gram-negative, coccobacillus South-East Asia Region (25-95%)	Pneumonia, UTI, septicaemia, Meningitis, Wound and surgical site infections	Beta-lactams, imipenem, meropenem, antipseudomonal agents, and fluoroquinolones,	Polymyxins, Polymyxins + carbapenem, minocycline, tigecycline, or rifampin,	[49-53]
<i>S. epidermidis</i> /gram-positive, cocci Africa (0-34%)	Bacteraemia and sepsis, catheter, implant, endocarditis and prostheses-related infection	Methicillin, penicillin, amoxicillin rifamycin, flouroquinolones, chloramphenicol, erythromycin, clindamycin, and sulfonamides	Vancomycin, vancomycin +/- (rifampin + gentamicin) daptomycin, linezolid, quinupristin-dalfopristin	[54-57]
<i>Burkholderia cepacia</i> / gram-negative, rod USA (0-40%)	Necrotizing pneumonia, peritonitis, liver abscesses	Chloramphenicol, Polymyxins, ciprofloxacin, moxifloxacin, colomycin, and aminoglycoside	Clinafloxacin, meropenem, ceftazidime and high-dose tobramycin (multiple combination is preferred)	[58-64]
<i>Neisseria gonorrhoeae</i> / gram-negative, diplococci European Region	Pelvic inflammatory disease, chronic pelvic pain, ectopic pregnancy, neonatal conjunctivitis, and infertility	Penicillin, ceftriaxone	Ciprofloxacin, spectinomycin and third-generation cephalosporins such as ceftriaxone and cefixime	[5,65,66]
<i>Clostridioides difficile</i> / gram-positive, rod shaped North America (0-10%)	Diarrhoea, colitis	Cefotaxime, levofloxacin, erythromycin, Cephalosporins	Rifampicin, meropenem, rifaximin	[67-70]

Supplementary Table 2. Principal AST methods, associated advantages and disadvantages (only examples are given).

Methods	Technique	Major advantages	Major disadvantages	References
Phenotypic	Disk diffusion	Simple, cost effective, flexible & well standardized	Lacks qualitative test & full- automation, risk of contamination, time consuming (minimum of 10 hours), no MIC determination, needs at least cells (~10 ⁵)	[71]
	Microdilution	Miniaturized, time feasible, allows MIC determination with multiple sampling	Inflexibility of drug selections available in standard commercial panels, time consuming (minimum of 10 hours), needs at least cells (~10 ⁵)	[72-75]
	Macrodilution	Simple, cost effective, allows MIC determination	Tedious sample preparation, risk of contamination, time consuming (minimum of 10 hours), needs at least cells (~10 ⁵) and expensive technique.	[76,77]
	Epsilon meter test	Detection of fastidious bacteria, easy to perform	High rate of false positive results, proper storage of Etest strip, (minimum of 10 hours), needs at least cells (~10 ⁵) Expensive (),	[78-82]
	MALDI-TOF MS	High throughput quick sample preparation automation possible	needs at least cells (~10 ⁵) expensive, different genetic markers	[83-85]
Genotypic	PCR	Quick (2-4 hours), simple and cost-effective, highly reproducible, high specificity, low sample requirement, Multiplexing,	Unknown parameters (mutation, G and C content, porosity of gel matrix, ionic strength and pH) can affect analysis, false positive results, expertise is required for analyses	[86,87]

		high sensitivity (picogram of nucleic acid)		
	DNA microarray s/chips	Multiplex detection, automated specific and high sensitivity (picogram of nucleic acid)	Labor intensive procedure to synthesize, purify, and store DNA before microarray fabrication, problem of cross hybridization	[88-90]
Automated	Vitek	Easy and fast data management, reproducible	Narrow antibiotic range requires more cells (~10 ⁵)	[91,92]
	Sensititre	Automated, rapid and simple operation	Narrow antibiotic range, limited panel capacity, expensive, needs at least cells (~10 ⁵)	[93-95]
	MicroScan Walkaway	Validate susceptibility with expert software system, accurate & data sharing	long incubation (minimum of 12 hours), expensive, needs at least cells (~10 ⁵)	[96-98]
	Micronaut	Direct susceptibility test	High false positive, needs at least cells (~10 ⁵)	[99]
	AvantagEst	Effortless, direct test	Discrepancies in results with some antibiotic combinations	[100-102]
Emerging AST	Optical	Real-time quantification of bacterial growth dynamics, time efficient (1-4 hours), morphological analysis, single cell detection	Requires microscope, cloning, genetic modifications may affect the growth dynamics	[103-105]
	Electrochemical	High sensitivity (10 ³ cells), economical, time efficient (1-4 hours)	Narrow temperature range Short life time of sensors interference with electroactive agents of sample	[106-108]

References

1. Reygaert, W. Methicillin-resistant Staphylococcus aureus (MRSA): identification and susceptibility testing techniques. *Clin Lab Sci* **2009**, *22*, 120-124.
2. Tong, S.Y.C.; Davis, J.S.; Eichenberger, E.; Holland, T.L.; Fowler, V.G. Staphylococcus aureus Infections: Epidemiology, Pathophysiology, Clinical Manifestations, and Management. *Clinical Microbiology Reviews* **2015**, *28*, 603-661, doi:10.1128/CMR.00134-14.
3. Guignard, B.; Entenza, J.M.; Moreillon, P. Beta-lactams against methicillin-resistant Staphylococcus aureus. *Curr Opin Pharmacol* **2005**, *5*, 479-489, doi:10.1016/j.coph.2005.06.002.
4. Scordo, K.A. Intravenous Antibiotics Used in the Treatment of Methicillin-Resistant Staphylococcus Aureus. *AACN Adv Crit Care* **2015**, *26*, 233-243, doi:10.1097/nci.000000000000095.
5. Organization, W.H. *Antimicrobial resistance: global report on surveillance*; World Health Organization: 2014.
6. Entenza, J.M.; Bétrisey, B.; Manuel, O.; Giddey, M.; Sakwinska, O.; Laurent, F.; Bizzini, A. Rapid Detection of Staphylococcus aureus Strains with Reduced Susceptibility to Vancomycin by Isothermal Microcalorimetry. *Journal of Clinical Microbiology* **2014**, *52*, 180-186, doi:10.1128/JCM.01820-13.
7. Liu, C.; Chambers, H.F. Staphylococcus aureus with Heterogeneous Resistance to Vancomycin: Epidemiology, Clinical Significance, and Critical Assessment of Diagnostic Methods. *Antimicrobial Agents and Chemotherapy* **2003**, *47*, 3040-3045, doi:10.1128/AAC.47.10.3040-3045.2003.
8. Hiramatsu, K.; Kayayama, Y.; Matsuo, M.; Aiba, Y.; Saito, M.; Hishinuma, T.; Iwamoto, A. Vancomycin-intermediate resistance in Staphylococcus aureus. *Journal of Global Antimicrobial Resistance* **2014**, *2*, 213-224, doi:<https://doi.org/10.1016/j.jgar.2014.04.006>.
9. Zhang, S.; Sun, X.; Chang, W.; Dai, Y.; Ma, X. Systematic Review and Meta-Analysis of the Epidemiology of Vancomycin-Intermediate and Heterogeneous Vancomycin-Intermediate Staphylococcus aureus Isolates. *PLoS One* **2015**, *10*, e0136082-e0136082, doi:10.1371/journal.pone.0136082.

10. Broaddus, V.C.; Light, R.W. Pleural effusion. In *Murray and Nadel's Textbook of Respiratory Medicine (Sixth Edition)*, Elsevier: 2016; pp. 1396-1424. e1310.
11. Murray, P.R.; Wold, A.D.; Schreck, C.A.; Washington Ja, I.I. Effects of selective media and atmosphere of incubation on the isolation of group A streptococci. *Journal of Clinical Microbiology* **1976**, *4*, 54-56.
12. Shulman, S.T.; Bisno, A.L.; Clegg, H.W.; Gerber, M.A.; Kaplan, E.L.; Lee, G.; Martin, J.M.; Van Beneden, C. Clinical practice guideline for the diagnosis and management of group A streptococcal pharyngitis: 2012 update by the Infectious Diseases Society of America. *Clin Infect Dis* **2012**, *55*, 1279-1282, doi:10.1093/cid/cis847.
13. Walker, M.J.; Barnett, T.C.; McArthur, J.D.; Cole, J.N.; Gillen, C.M.; Henningham, A.; Sriprakash, K.S.; Sanderson-Smith, M.L.; Nizet, V. Disease Manifestations and Pathogenic Mechanisms of Group A Streptococcus. *Clinical Microbiology Reviews* **2014**, *27*, 264-301, doi:10.1128/CMR.00101-13.
14. Efstratiou, A.; Lamagni, T. Epidemiology of Streptococcus pyogenes. In *Streptococcus pyogenes: basic biology to clinical manifestations [Internet]*, University of Oklahoma Health Sciences Center: 2017.
15. Blaschke, A.J. Interpreting Assays for the Detection of Streptococcus pneumoniae. *Clinical Infectious Diseases: An Official Publication of the Infectious Diseases Society of America* **2011**, *52*, S331-S337, doi:10.1093/cid/cir048.
16. Pikiš, A.; Campos, J.M.; Rodriguez, W.J.; Keith, J.M. Optochin Resistance in Streptococcus pneumoniae: Mechanism, Significance, and Clinical Implications. *The Journal of Infectious Diseases* **2001**, *184*, 582-590, doi:10.1086/322803.
17. Kaplan, S.L.; Mason, E.O., Jr. Management of infections due to antibiotic-resistant Streptococcus pneumoniae. *Clin Microbiol Rev* **1998**, *11*, 628-644.
18. Molina, F.; López-Acedo, E.; Tabla, R.; Roa, I.; Gómez, A.; Rebollo, J.E. Improved detection of Escherichia coli and coliform bacteria by multiplex PCR. *BMC Biotechnology* **2015**, *15*, 48, doi:10.1186/s12896-015-0168-2.
19. Sackmann, E.K.; Fulton, A.L.; Beebe, D.J. The present and future role of microfluidics in biomedical research. *Nature* **2014**, *507*, 181-189, doi:10.1038/nature13118.
20. Rodriguez-Bano, J.; Alcalá, J.C.; Cisneros, J.M.; Grill, F.; Oliver, A.; Horcajada, J.P.; Tortola, T.; Mirelis, B.; Navarro, G.; Cuenca, M., et al. Community infections caused by extended-spectrum beta-lactamase-producing Escherichia coli. *Arch Intern Med* **2008**, *168*, 1897-1902, doi:10.1001/archinte.168.17.1897.
21. Morgand, M.; Vimont, S.; Bleibtreu, A.; Boyd, A.; Thien, H.V.; Zahar, J.-R.; Denamur, E.; Arlet, G. Extended-spectrum beta-lactamase-producing Escherichia coli infections in children: Are community-acquired strains different from nosocomial strains? *International Journal of Medical Microbiology* **2014**, *304*, 970-976, doi:https://doi.org/10.1016/j.ijmm.2014.06.003.
22. Lim, J.Y.; Yoon, J.W.; Hovde, C.J. A Brief Overview of Escherichia coli O157:H7 and Its Plasmid O157. *Journal of microbiology and biotechnology* **2010**, *20*, 5-14.
23. Ch. Schröder, U.; Beileites, C.; Assmann, C.; Glaser, U.; Hübner, U.; Pfister, W.; Fritzsche, W.; Popp, J.; Neugebauer, U. Detection of vancomycin resistances in enterococci within 3 ½ hours. *Scientific Reports* **2015**, *5*, 8217, doi:10.1038/srep08217 <https://www.nature.com/articles/srep08217#supplementary-information>.
24. Hrabak, J.; Chudackova, E.; Walkova, R. Matrix-assisted laser desorption ionization-time of flight (maldi-tof) mass spectrometry for detection of antibiotic resistance mechanisms: from research to routine diagnosis. *Clin Microbiol Rev* **2013**, *26*, 103-114, doi:10.1128/cmr.00058-12.
25. Lupo, A.; Papp-Wallace, K.M.; Sendi, P.; Bonomo, R.A.; Endimiani, A. Non-phenotypic tests to detect and characterize antibiotic resistance mechanisms in Enterobacteriaceae. *Diagn Microbiol Infect Dis* **2013**, *77*, 179-194, doi:10.1016/j.diagmicrobio.2013.06.001.

26. Pulido, M.R.; Garcia-Quintanilla, M.; Martin-Pena, R.; Cisneros, J.M.; McConnell, M.J. Progress on the development of rapid methods for antimicrobial susceptibility testing. *J Antimicrob Chemother* **2013**, *68*, 2710-2717, doi:10.1093/jac/dkt253.
27. Wiegand, I.; Hilpert, K.; Hancock, R.E. Agar and broth dilution methods to determine the minimal inhibitory concentration (MIC) of antimicrobial substances. *Nat Protoc* **2008**, *3*, 163-175, doi:10.1038/nprot.2007.521.
28. Chewning, J.H. Vancomycin-resistant Enterococcus faecium Bacteremia Successfully Treated With High-dose Ampicillin-Sulbactam in a Pediatric Patient After Hematopoietic Stem Cell Transplantation. *Journal of Pediatric Hematology/Oncology* **2011**, *33*, 401, doi:10.1097/MPH.0b013e31820db7eb.
29. O'Driscoll, T.; Crank, C.W. Vancomycin-resistant enterococcal infections: epidemiology, clinical manifestations, and optimal management. *Infection and drug resistance* **2015**, *8*, 217-230, doi:10.2147/IDR.S54125.
30. Kohler, P.; Eshaghi, A.; Kim, H.C.; Plevneshi, A.; Green, K.; Willey, B.M.; McGeer, A.; Patel, S.N. Prevalence of vancomycin-variable Enterococcus faecium (VVE) among vanA-positive sterile site isolates and patient factors associated with VVE bacteremia. *PLoS One* **2018**, *13*, e0193926, doi:10.1371/journal.pone.0193926.
31. Xu, J.; Moore, J.E.; Murphy, P.G.; Millar, B.C.; Elborn, J.S. Early detection of Pseudomonas aeruginosa – comparison of conventional versus molecular (PCR) detection directly from adult patients with cystic fibrosis (CF). *Annals of Clinical Microbiology and Antimicrobials* **2004**, *3*, 21-21, doi:10.1186/1476-0711-3-21.
32. Khan, A.A.; Cerniglia, C.E. Detection of Pseudomonas aeruginosa from clinical and environmental samples by amplification of the exotoxin A gene using PCR. *Applied and Environmental Microbiology* **1994**, *60*, 3739-3745.
33. Counts, G.W.; Schwartz, R.W.; Ulness, B.K.; Hamilton, D.J.; Rosok, M.J.; Cunningham, M.D.; Tam, M.R.; Darveau, R.P. Evaluation of an immunofluorescent-antibody test for rapid identification of Pseudomonas aeruginosa in blood cultures. *J Clin Microbiol* **1988**, *26*, 1161-1165.
34. Hachem, R.Y.; Chemaly, R.F.; Ahmar, C.A.; Jiang, Y.; Boktour, M.R.; Rjaili, G.A.; Bodey, G.P.; Raad, I.I. Colistin Is Effective in Treatment of Infections Caused by Multidrug-Resistant Pseudomonas aeruginosa in Cancer Patients. *Antimicrobial Agents and Chemotherapy* **2007**, *51*, 1905-1911, doi:10.1128/AAC.01015-06.
35. Tan, J.S.; File, T.M., Jr. Antipseudomonal penicillins. *Med Clin North Am* **1995**, *79*, 679-693.
36. Palavutitotai, N.; Jitmuang, A.; Tongchai, S.; Kiratisin, P.; Angkasekwinai, N. Epidemiology and risk factors of extensively drug-resistant Pseudomonas aeruginosa infections. *PLoS One* **2018**, *13*, e0193431-e0193431, doi:10.1371/journal.pone.0193431.
37. Khuntayaporn, P.; Montakantikul, P.; Santanirand, P.; Kiratisin, P.; Chomnawang, M.T. Molecular investigation of carbapenem resistance among multidrug-resistant Pseudomonas aeruginosa isolated clinically in Thailand. *Microbiol. Immunol.* **2013**, *57*, 170-178, doi:10.1111/1348-0421.12021.
38. Gupta, A.; Ampofo, K.; Rubenstein, D.; Saiman, L. Extended Spectrum β Lactamase-producing Klebsiella pneumoniae Infections: a Review of the Literature. *Journal Of Perinatology* **2003**, *23*, 439, doi:10.1038/sj.jp.7210973.
39. Sekowska, A.; Janicka, G.; Klyszejko, C.; Wojda, M.; Wroblewski, M.; Szymankiewicz, M. Resistance of Klebsiella pneumoniae strains producing and not producing ESBL (extended-spectrum beta-lactamase) type enzymes to selected non-beta-lactam antibiotics. *Med Sci Monit* **2002**, *8*, Br100-104.
40. Lee, C.H.; Su, L.H.; Tang, Y.F.; Liu, J.W. Treatment of ESBL-producing Klebsiella pneumoniae bacteraemia with carbapenems or flomoxef: a retrospective study and laboratory analysis of the isolates. *J Antimicrob Chemother* **2006**, *58*, 1074-1077, doi:10.1093/jac/dkl381.

41. Metri, B.C.; Jyothi, P.; Peerapur, B.V. Detection of ESBL in E.coli and K. pneumoniae isolated from urinary tract infection. *Indian Journal of Nephrology* **2012**, *22*, 401-402, doi:10.4103/0971-4065.103919.
42. Ghasemi, Y.; Archin, T.; Kargar, M.; Mohkam, M. A simple multiplex PCR for assessing prevalence of extended-spectrum β -lactamases producing *Klebsiella pneumoniae* in Intensive Care Units of a referral hospital in Shiraz, Iran. *Asian Pacific Journal of Tropical Medicine* **2013**, *6*, 703-708, doi:https://doi.org/10.1016/S1995-7645(13)60122-4.
43. Goble, M.; Iseman, M.D.; Madsen, L.A.; Waite, D.; Ackerson, L.; Horsburgh, C.R.J. Treatment of 171 Patients with Pulmonary Tuberculosis Resistant to Isoniazid and Rifampin. *New England Journal of Medicine* **1993**, *328*, 527-532, doi:10.1056/nejm199302253280802.
44. Ziganshina, L.E.; Vigel, A.A.; Squire, S.B. Fluoroquinolones for treating tuberculosis. *Cochrane Database of Systematic Reviews* **2005**, 10.1002/14651858.CD004795.pub2, doi:10.1002/14651858.CD004795.pub2.
45. Mitnick, C.D.; Appleton, S.C.; Shin, S.S. Epidemiology and Treatment of Multidrug Resistant Tuberculosis. *Seminars in respiratory and critical care medicine* **2008**, *29*, 499-524, doi:10.1055/s-0028-1085702.
46. Pigrau-Serrallach, C.; Rodríguez-Pardo, D. Bone and joint tuberculosis. *European Spine Journal* **2013**, *22*, 556-566, doi:10.1007/s00586-012-2331-y.
47. Choi, W.; Lee, J.; Cho, E.; Jung, G.Y. Accurate and effective multidrug-resistant *Mycobacterium tuberculosis* detection method using gap-filling ligation coupled with high-resolution capillary electrophoresis-based single strand conformation polymorphism. *Scientific Reports* **2017**, *7*, 46090, doi:10.1038/srep46090.
48. Ormerod, L.P. Multidrug-resistant tuberculosis (MDR-TB): epidemiology, prevention and treatment. *Br. Med. Bull.* **2005**, *73-74*, 17-24, doi:10.1093/bmb/ldh047.
49. Howard, A.; O'Donoghue, M.; Feeney, A.; Sleator, R.D. *Acinetobacter baumannii*: An emerging opportunistic pathogen. *Virulence* **2012**, *3*, 243-250, doi:10.4161/viru.19700.
50. Guerrero, D.M.; Perez, F.; Conger, N.G.; Solomkin, J.S.; Adams, M.D.; Rather, P.N.; Bonomo, R.A. *Acinetobacter baumannii*-Associated Skin and Soft Tissue Infections: Recognizing a Broadening Spectrum of Disease. *Surgical Infections* **2010**, *11*, 49-57, doi:10.1089/sur.2009.022.
51. Li, P.; Niu, W.k.; Li, H.; Lei, H.; Liu, W.; Zhao, X.; Guo, L.; Zou, D.; Yuan, X.; Liu, H., et al. Rapid detection of *Acinetobacter baumannii* and molecular epidemiology of carbapenem-resistant *A. baumannii* in two comprehensive hospitals of Beijing, China. *Frontiers in Microbiology* **2015**, *6*, doi:10.3389/fmicb.2015.00997.
52. Turton, J.F.; Woodford, N.; Glover, J.; Yarde, S.; Kaufmann, M.E.; Pitt, T.L. Identification of *Acinetobacter baumannii* by Detection of the bla(OXA-51-like) Carbapenemase Gene Intrinsic to This Species. *Journal of Clinical Microbiology* **2006**, *44*, 2974-2976, doi:10.1128/JCM.01021-06.
53. Suwantararat, N.; Carroll, K.C. Epidemiology and molecular characterization of multidrug-resistant Gram-negative bacteria in Southeast Asia. *Antimicrobial resistance and infection control* **2016**, *5*, 15-15, doi:10.1186/s13756-016-0115-6.
54. Rogers, K.L.; Fey, P.D.; Rupp, M.E. Coagulase-negative staphylococcal infections. *Infect Dis Clin North Am* **2009**, *23*, 73-98, doi:10.1016/j.idc.2008.10.001.
55. Otto, M. *Staphylococcus epidermidis* – the “accidental” pathogen. *Nature reviews. Microbiology* **2009**, *7*, 555-567, doi:10.1038/nrmicro2182.
56. Ikeda, Y.; Ohara-Nemoto, Y.; Kimura, S.; Ishibashi, K.; Kikuchi, K. PCR-based identification of *Staphylococcus epidermidis* targeting *gseA* encoding the glutamic-acid-specific protease. *Canadian Journal of Microbiology* **2004**, *50*, 493-498, doi:10.1139/w04-055.
57. Chessa, D.; Ganau, G.; Mazzarello, V. An overview of *Staphylococcus epidermidis* and *Staphylococcus aureus* with a focus on developing countries. *Journal of infection in developing countries* **2015**, *9*, 547-550, doi:10.3855/jidc.6923.

58. Poole, K. Efflux-Mediated Resistance to Fluoroquinolones in Gram-Negative Bacteria. *Antimicrobial Agents and Chemotherapy* **2000**, *44*, 2233-2241.
59. Zhang, L.; Li, X.-Z.; Poole, K. Fluoroquinolone susceptibilities of efflux-mediated multidrug-resistant *Pseudomonas aeruginosa*, *Stenotrophomonas maltophilia* and *Burkholderia cepacia*. *Journal of Antimicrobial Chemotherapy* **2001**, *48*, 549-552, doi:10.1093/jac/48.4.549.
60. Horsley, A.; Jones, A.M.; Lord, R. Antibiotic treatment for *Burkholderia cepacia* complex in people with cystic fibrosis experiencing a pulmonary exacerbation. *Cochrane Database of Systematic Reviews* **2016**, 10.1002/14651858.CD009529.pub3, doi:10.1002/14651858.CD009529.pub3.
61. Aaron, S.D.; Ferris, W.; Henry, D.A.; Speert, D.P.; Macdonald, N.E. Multiple combination bactericidal antibiotic testing for patients with cystic fibrosis infected with *Burkholderia cepacia*. *Am J Respir Crit Care Med* **2000**, *161*, 1206-1212, doi:10.1164/ajrccm.161.4.9907147.
62. Mukhopadhyay, C.; Bhargava, A.; Ayyagari, A. Two novel clinical presentations of *Burkholderia cepacia* infection. *J Clin Microbiol* **2004**, *42*, 3904-3905, doi:10.1128/jcm.42.8.3904-3905.2004.
63. McDowell, A.; Mahenthalingam, E.; Moore, J.E.; Dunbar, K.E.; Webb, A.K.; Dodd, M.E.; Martin, S.L.; Millar, B.C.; Scott, C.J.; Crowe, M., et al. PCR-based detection and identification of *Burkholderia cepacia* complex pathogens in sputum from cystic fibrosis patients. *J Clin Microbiol* **2001**, *39*, 4247-4255, doi:10.1128/jcm.39.12.4247-4255.2001.
64. Spencer, R.C. The emergence of epidemic, multiple-antibiotic-resistant *Stenotrophomonas* (*Xanthomonas*) *maltophilia* and *Burkholderia* (*Pseudomonas*) *cepacia*. *J. Hosp. Infect.* **1995**, *30*, 453-464, doi:https://doi.org/10.1016/0195-6701(95)90049-7.
65. Geraats-Peters, C.W.M.; Brouwers, M.; Schneeberger, P.M.; van der Zanden, A.G.M.; Bruisten, S.M.; Weers-Pothoff, G.; Boel, C.H.E.; van den Brule, A.J.C.; Harmsen, H.G.; Hermans, M.H.A. Specific and Sensitive Detection of *Neisseria gonorrhoeae* in Clinical Specimens by Real-Time PCR. *Journal of Clinical Microbiology* **2005**, *43*, 5653-5659, doi:10.1128/JCM.43.11.5653-5659.2005.
66. Ng, L.-K.; Martin, I.E. The laboratory diagnosis of *Neisseria gonorrhoeae*. *The Canadian Journal of Infectious Diseases & Medical Microbiology* **2005**, *16*, 15-25.
67. Yoldaş, Ö.; Altındiş, M.; Cufalı, D.; Aşık, G.; Keşli, R. A Diagnostic Algorithm for the Detection of *Clostridium difficile*-Associated Diarrhea. *Balkan Medical Journal* **2016**, *33*, 80-86, doi:10.5152/balkanmedj.2015.15159.
68. Huang, H.; Weintraub, A.; Fang, H.; Nord, C.E. Antimicrobial resistance in *Clostridium difficile*. *International Journal of Antimicrobial Agents* **2009**, *34*, 516-522, doi:https://doi.org/10.1016/j.ijantimicag.2009.09.012.
69. Spigaglia, P. Recent advances in the understanding of antibiotic resistance in *Clostridium difficile* infection. *Therapeutic Advances in Infectious Disease* **2016**, *3*, 23-42, doi:10.1177/2049936115622891.
70. Balsells, E.; Shi, T.; Leese, C.; Lyell, I.; Burrows, J.; Wiuff, C.; Campbell, H.; Kyaw, M.H.; Nair, H. Global burden of *Clostridium difficile* infections: a systematic review and meta-analysis. *Journal of global health* **2019**, *9*, 010407-010407, doi:10.7189/jogh.09.010407.
71. Delignette-Muller, M.; Flandrois, J. An accurate diffusion method for determining bacterial sensitivity to antibiotics. *Journal of antimicrobial chemotherapy* **1994**, *34*, 73-81.
72. McDermott, P.F.; Bodeis-Jones, S.M.; Fritsche, T.R.; Jones, R.N.; Walker, R.D. Broth microdilution susceptibility testing of *Campylobacter jejuni* and the determination of quality control ranges for fourteen antimicrobial agents. *Journal of clinical microbiology* **2005**, *43*, 6136-6138.
73. Murray, C.K.; Hospenthal, D.R. Broth microdilution susceptibility testing for *Leptospira* spp. *Antimicrobial agents and chemotherapy* **2004**, *48*, 1548-1552.

74. Miller, R.A.; Walker, R.D.; Carson, J.; Coles, M.; Coyne, R.; Dalsgaard, I.; Giesecker, C.; Hsu, H.M.; Mathers, J.J.; Papapetropoulou, M., et al. Standardization of a broth microdilution susceptibility testing method to determine minimum inhibitory concentrations of aquatic bacteria. *Diseases of aquatic organisms* **2005**, *64*, 211-222, doi:10.3354/dao064211.
75. Waites, K.B.; Duffy, L.B.; Bébéar, C.M.; Matlow, A.; Talkington, D.F.; Kenny, G.E.; Totten, P.A.; Bade, D.J.; Zheng, X.; Davidson, M.K. Standardized methods and quality control limits for agar and broth microdilution susceptibility testing of *Mycoplasma pneumoniae*, *Mycoplasma hominis*, and *Ureaplasma urealyticum*. *Journal of clinical microbiology* **2012**, JCM. 01439-01412.
76. Tomida, J.; Oumi, A.; Okamoto, T.; Morita, Y.; Okayama, A.; Misawa, N.; Hayashi, T.; Akaike, T.; Kawamura, Y. Comparative evaluation of agar dilution and broth microdilution methods for antibiotic susceptibility testing of *Helicobacter cinaedi*. *Microbiology and immunology* **2013**, *57*, 353-358, doi:10.1111/1348-0421.12044.
77. Siddiqi, S.H.; Heifets, L.B.; Cynamon, M.H.; Hooper, N.M.; Laszlo, A.; Libonati, J.P.; Lindholm-Levy, P.J.; Pearson, N. Rapid broth macrodilution method for determination of MICs for *Mycobacterium avium* isolates. *J Clin Microbiol* **1993**, *31*, 2332-2338.
78. Glupczynski, Y.; Labbe, M.; Hansen, W.; Crokaert, F.; Yourassowsky, E. Evaluation of the E test for quantitative antimicrobial susceptibility testing of *Helicobacter pylori*. *Journal of Clinical Microbiology* **1991**, *29*, 2072-2075.
79. Brown, D.F.; Brown, L. Evaluation of the E test, a novel method of quantifying antimicrobial activity. *The Journal of antimicrobial chemotherapy* **1991**, *27*, 185-190.
80. Baker, C.N. The E-Test and *Campylobacter jejuni*. *Diagnostic microbiology and infectious disease* **1992**, *15*, 469-472.
81. Schulz, J.; Sahm, D. Reliability of the E test for detection of ampicillin, vancomycin, and high-level aminoglycoside resistance in *Enterococcus* spp. *Journal of clinical microbiology* **1993**, *31*, 3336-3339.
82. Cantón, R.; Livermore, D.M.; Morosini, M.I.; Díaz-Regañón, J.; Rossolini, G.M.; Group, P.S.; Group, P.S. Etest® versus broth microdilution for ceftaroline MIC determination with *Staphylococcus aureus*: results from PREMIUM, a European multicentre study. *Journal of Antimicrobial Chemotherapy* **2016**, dkw442.
83. Clark, A.E.; Kaleta, E.J.; Arora, A.; Wolk, D.M. Matrix-assisted laser desorption ionization–time of flight mass spectrometry: a fundamental shift in the routine practice of clinical microbiology. *Clinical microbiology reviews* **2013**, *26*, 547-603.
84. Singhal, N.; Kumar, M.; Kanaujia, P.K.; Viridi, J.S. MALDI-TOF mass spectrometry: an emerging technology for microbial identification and diagnosis. *Frontiers in Microbiology* **2015**, *6*, doi:10.3389/fmicb.2015.00791.
85. Tré-Hardy, M.; Lambert, B.; Despas, N.; Bressant, F.; Laurenzano, C.; Rodriguez-Villalobos, H.; Verroken, A. MALDI-TOF MS identification and antimicrobial susceptibility testing directly from positive enrichment broth. *Journal of Microbiological Methods* **2017**, *141*, 32-34, doi:https://doi.org/10.1016/j.mimet.2017.07.012.
86. Strommenger, B.; Kettlitz, C.; Werner, G.; Witte, W. Multiplex PCR assay for simultaneous detection of nine clinically relevant antibiotic resistance genes in *Staphylococcus aureus*. *J Clin Microbiol* **2003**, *41*, 4089-4094.
87. Cockerill, F.R. Genetic methods for assessing antimicrobial resistance. *Antimicrobial agents and chemotherapy* **1999**, *43*, 199-212.
88. Strommenger, B.; Schmidt, C.; Werner, G.; Roessle-Lorch, B.; Bachmann, T.T.; Witte, W. DNA microarray for the detection of therapeutically relevant antibiotic resistance determinants in clinical isolates of *Staphylococcus aureus*. *Molecular and cellular probes* **2007**, *21*, 161-170, doi:10.1016/j.mcp.2006.10.003.
89. Miller, M.B.; Tang, Y.-W. Basic Concepts of Microarrays and Potential Applications in Clinical Microbiology. *Clinical Microbiology Reviews* **2009**, *22*, 611-633, doi:10.1128/CMR.00019-09.

90. Lu, N.; Hu, Y.; Zhu, L.; Yang, X.; Yin, Y.; Lei, F.; Zhu, Y.; Du, Q.; Wang, X.; Meng, Z., et al. DNA microarray analysis reveals that antibiotic resistance-gene diversity in human gut microbiota is age related. *Scientific Reports* **2014**, *4*, 4302, doi:10.1038/srep04302

<https://www.nature.com/articles/srep04302#supplementary-information>.

91. Bobenchik, A.M.; Hindler, J.A.; Giltner, C.L.; Saeki, S.; Humphries, R.M. Performance of Vitek 2 for Antimicrobial Susceptibility Testing of Staphylococcus spp. and Enterococcus spp. *Journal of Clinical Microbiology* **2014**, *52*, 392-397, doi:10.1128/JCM.02432-13.
92. van Belkum, A.; Dunne, W.M. Next-generation antimicrobial susceptibility testing. *Journal of clinical microbiology* **2013**, *51*, 2018-2024.
93. Pykett, A.H. An assessment of the Sensititre method for determining minimum inhibitory and bactericidal concentrations. *Journal of clinical pathology* **1978**, *31*, 536-538.
94. Doern, G.; Staneck, J.; Needham, C.; Tubert, T. Sensititre autoreader for same-day breakpoint broth microdilution susceptibility testing of members of the family Enterobacteriaceae. *Journal of clinical microbiology* **1987**, *25*, 1481-1485.
95. Staneck, J.L.; Allen, S.D.; Harris, E.E.; Tilton, R.C. Rapid MIC testing with the sensititre autoreader. *Journal of clinical microbiology* **1988**, *26*, 1-7.
96. Richter, S.S.; Ferraro, M.J. Susceptibility testing instrumentation and computerized expert systems for data analysis and interpretation. In *Manual of Clinical Microbiology, 10th Edition*, American Society of Microbiology: ASM Press, P.O. Box 605, Herndon, VA, USA 2011; pp. 1144-1154.
97. Jang, W.; Park, Y.-J.; Park, K.G.; Yu, J. Evaluation of MicroScan WalkAway and Vitek 2 for determination of the susceptibility of extended-spectrum β -lactamase-producing *Escherichia coli* and *Klebsiella pneumoniae* isolates to cefepime, cefotaxime and ceftazidime. *Journal of Antimicrobial Chemotherapy* **2013**, *68*, 2282-2285, doi:10.1093/jac/dkt172.
98. Burns, J.L.; Saiman, L.; Whittier, S.; Krzewinski, J.; Liu, Z.; Larone, D.; Marshall, S.A.; Jones, R.N. Comparison of two commercial systems (Vitek and MicroScan-WalkAway) for antimicrobial susceptibility testing of *Pseudomonas aeruginosa* isolates from cystic fibrosis patients. *Diagnostic microbiology and infectious disease* **2001**, *39*, 257-260.
99. Wellinghausen, N.; Pietzcker, T.; Poppert, S.; Belak, S.; Fieser, N.; Bartel, M.; Essig, A. Evaluation of the Merlin MICRONAUT System for Rapid Direct Susceptibility Testing of Gram-Positive Cocci and Gram-Negative Bacilli from Positive Blood Cultures. *Journal of Clinical Microbiology* **2007**, *45*, 789-795, doi:10.1128/jcm.01856-06.
100. Wright, D.N.; Matsen, J.M.; DiPersio, J.R.; Kirk, M.; Saxon, B.; Ficorilli, S.M.; Spencer, H.J. Evaluation of four newer antimicrobial agents in the Avantage susceptibility test system. *Journal of Clinical Microbiology* **1989**, *27*, 2381-2383.
101. Nolte, F.S.; Contestable, P.B.; Lincalis, D.; Punsalang Jr, A. Rapid, direct antibiotic susceptibility testing of blood culture isolates using the Abbott advantage[®] system. *American journal of clinical pathology* **1986**, *86*, 665-669.
102. Sriboonsong, S.; Boonchoo, L. Automatic antimicrobial susceptibility testing system. In *Proceedings of World Congress on Medical Physics and Biomedical Engineering 2006*; pp. 2501-2504.
103. Golchin, S.A.; Stratford, J.; Curry, R.J.; McFadden, J. A microfluidic system for long-term time-lapse microscopy studies of mycobacteria. *Tuberculosis* **2012**, *92*, 489-496.
104. Mohan, R.; Mukherjee, A.; Sevgen, S.E.; Sanpitakserree, C.; Lee, J.; Schroeder, C.M.; Kenis, P.J.A. A multiplexed microfluidic platform for rapid antibiotic susceptibility testing. *Biosensors and Bioelectronics* **2013**, *49*, 118-125.
105. Mohan, R.; Sanpitakserree, C.; Desai, A.V.; Sevgen, S.E.; Schroeder, C.M.; Kenis, P.J.A. A microfluidic approach to study the effect of bacterial interactions on antimicrobial susceptibility in polymicrobial cultures. *RSC Adv.* **2015**, *5*, 35211-35223, doi:10.1039/C5RA04092B.

106. Webster, T.A.; Sismaet, H.J.; Chan, I.p.J.; Goluch, E.D. Electrochemically monitoring the antibiotic susceptibility of *Pseudomonas aeruginosa* biofilms. *Analyst* **2015**, *140*, 7195-7201, doi:10.1039/C5AN01358E.
107. Baker, C.A.; Duong, C.T.; Grimley, A.; Roper, M.G. Recent advances in microfluidic detection systems. *Bioanalysis* **2009**, *1*, 967-975, doi:10.4155/bio.09.86.
108. Wang, Y.; Xu, H.; Zhang, J.; Li, G. Electrochemical Sensors for Clinic Analysis. *Sensors (Basel, Switzerland)* **2008**, *8*, 2043-2081.